

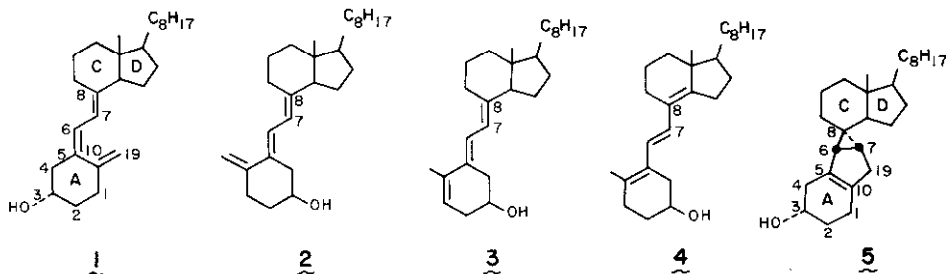
THE MASS SPECTRA OF SUPRASTEROL<sub>3</sub>-II, VITAMIN D<sub>3</sub> AND RELATED MOLECULES<sup>1</sup>

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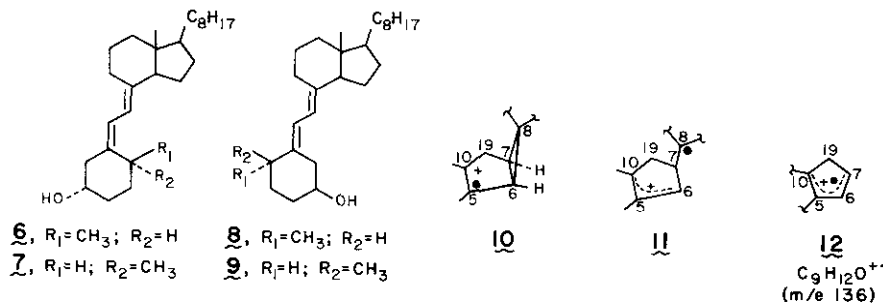
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The mass spectra of vitamin D<sub>3</sub> (1)<sup>2</sup> and 5,6-trans-vitamin D<sub>3</sub> (2)<sup>3</sup> are almost identical and are uniquely characterized by the appearance of a base peak at m/e 136 with the second most intense ion appearing at m/e 118 (base peak minus water). It is of some mechanistic concern that the m/e 136 peak (C<sub>9</sub>H<sub>12</sub>O<sup>+</sup>) for both substances is due to the A-ring plus HC-7, HC-6 and H<sub>2</sub>C-19, which would formally require the energetically unfavorable electron impact induced cleavage across the C-7/C-8 carbon-carbon double bond.<sup>4</sup> Neither isovitamin D<sub>3</sub> (3)<sup>5</sup> nor isotachysterol<sub>3</sub> (4),<sup>5</sup> double bond shifted structural isomers of 1, exhibit similar mass spectral fragmentation patterns particularly in regard to significant peaks at m/e 136 and 118. In point of fact the observation of these peaks, usually as the two most intense ions, appears to be general for metabolites and analogs of 1 and 2 possessing the same stereostructural triene arrangement.<sup>2,3,5a,6</sup>



Investigations of fragmentation reactions occurring in the mass spectrometer have revealed processes analogous to those observed both in thermal and in photochemical reactions.<sup>7</sup> Studies have been reported from a theoretical<sup>8</sup> and experimental<sup>4,7,9</sup> viewpoint concerning whether ground or electronically excited states are involved in reactions induced by electron impact. It is known that 1 and 2 are not interconverted by heat,<sup>10</sup> but that 2 reverts to 1 upon UV irradiation.<sup>11</sup> It is also known that upon irradiation of 1, the major photoproduct (62%) is the bicyclo[3.1.0]hexene suprasterol<sub>3</sub>-II (5).<sup>11b,12</sup> Because of the sometimes ob-

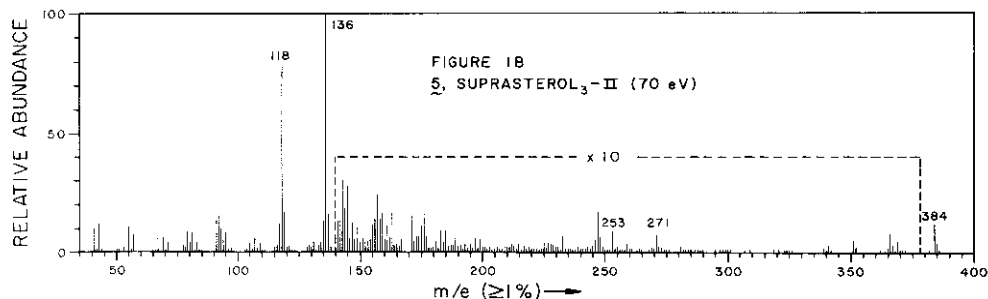
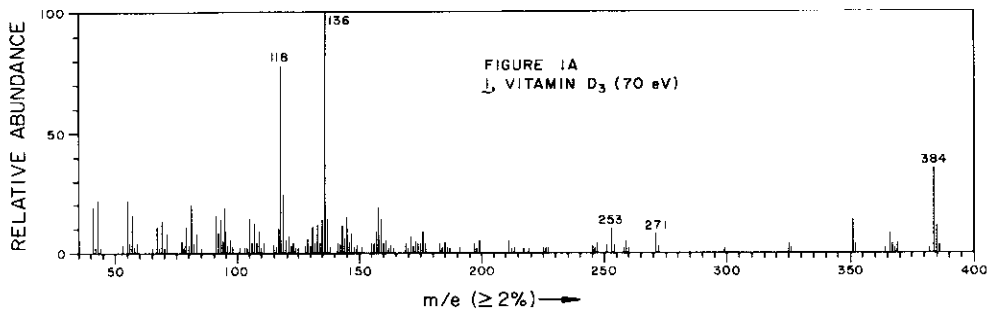
served parallel between photochemical and electron impact induced pathways,<sup>13</sup> it seemed attractive to investigate the mass spectral behavior of vitamin D<sub>3</sub> related valence isomers. We report herein a comparison of the mass spectrum of 5 with that of 1, and we also describe briefly the mass spectra of the four possible 10, 19-dihydro reduction products (6 - 9)<sup>14</sup> of 1 and 2 for further comparison.



The mass spectra (70 eV) of 1 and 5 are given in Figures 1A and 1B respectively. The spectra of 1, 5, and 2 (not shown)<sup>15</sup> are dominated by the m/e 136 (base) and 118 (78%, 79% and 76%, respectively) peaks and all three spectra exhibit metastables at m/e 48.2 and 102.4 attributable to the processes m/e 384 → 136 and 136 → 118, respectively. Slow scans at 70 and 20 eV showed that for the three compounds the broad metastable peaks at m/e 48.2 are virtually superimposable, and the same was observed for the narrow metastable peaks at m/e 102.4.<sup>16a</sup> Moreover, the spectra of 1 and 2 are nearly identical. Below m/e 140, the spectra of 1 and 5 are also almost identical; although at higher masses peaks due to 5 are less intense than those for 1, similarities are apparent at a tenfold expanded intensity scale (Figure 1B). Fragments at m/e 271 and 253, characteristic of the loss of the side chain and 271-H<sub>2</sub>O, respectively, previously mentioned for 1<sup>2</sup> and 2,<sup>3</sup> are also observed for 5. Abundance ratios for 1 and 5 of competitive metastable ion fragmentations in the first field-free region (so called HV scans<sup>16a,b</sup>) at 70 and 20 eV indicate that although some fragments arise from different precursors, those at m/e 369, 271, 259, 253, 211 (and 136, from the metastable peak shapes) appear to be generated from precursors having a common structure.

In view of the near-identity of the spectra observed for 1 and 2,<sup>3,15</sup> we also examined the spectra of 6 - 9 which differ from one another only in their C-10 and/or their Δ<sup>5</sup> configurations.<sup>17</sup> The four stereoisomers exhibit qualitative similarities:<sup>18</sup> m/e 386 (parent), 302, 301, 273, 259, 255, 247 and groups of peaks at 121,119/ 107,105/ 95,93,91/ 81,79/ 71,69,67/ 57,55/ and 43. The distribution of intensities differs widely, however. Most significantly, a peak resulting from C-7/C-8 cleavage (m/e 138) is absent (<0.5%) for all four compounds. These observations complement the earlier results in which neither 3 nor 4 exhibited significant C-7/C-8 scission fragments upon electron impact.<sup>5</sup>

As regards a possible mechanistic pathway through which 5 traverses to give the m/e 136 species (presumably 12), obvious choices for intermediates include



10 and 11. The mass spectral results described above indicate that the major decomposition path of the molecular ions of 1, 2 and 5 proceeds via a common intermediate structure or set of structures (possibly 10 and/or 11) to the same m/e 136 species (12). Since the three isomers are photochemically but not thermally interconnected, this finding suggests a parallel between the mass spectrometric and photolytic reaction paths.<sup>19</sup> The intermediacy of structures like 10 and/or 11 nicely accounts for the formal cleavage across the C-7/C-8 bond in 1 and 2, while no such scission occurs in compounds 3, 4 and 6-9.

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- (a) Paper XI (University of California, Riverside) in the series, Studies on Vitamin D (Calciferol) and Its Analogs. For Paper X, see R. L. Johnson, S. C. Carey, A. W. Norman and W. H. Okamura, *J. Medicinal Chem.*, in press; (b) Paper XXV (Leiden University) in the series, Studies on Vitamin D and Related Compounds. For Paper XXIV, see F. Boomsma, H. J. C. Jacobs, E. Havinga and A. van der Gen, *Tetrahedron Lett.*, 427 (1975).
- J. W. Blunt, H. F. DeLuca and H. K. Schnoes, *Biochem.*, **7**, 3317 (1968).
- M. F. Holick, M. Garabedian and H. F. DeLuca, *ibid.*, **11**, 2715 (1972).

4. For similar reasoning, see A. L. Burlingame, C. Fenselau, W. J. Richter, W. G. Dauben, G. W. Shaffer and N. C. Vietmeyer, J. Am. Chem. Soc., 89, 3346 (1967).
5. (a) D. E. M. Lawson, D. R. Fraser, E. Kodicek, H. R. Morris and D. H. Williams, Nature, 230, 228 (1971); (b) M. F. Holick, H. F. DeLuca, P. M. Kasten and M. B. Kōřycka, Science, 180, 964 (1973).
6. For examples, see (a) M. F. Holick, H. K. Schnoes and H. F. DeLuca, Proc. Nat. Acad. Sci. U.S.A., 68, 803 (1971); (b) A. W. Norman, J. F. Myrtle, R. J. Midgett, H. G. Nowicki, V. Williams and G. Popjak, Science, 173, 51 (1971); (c) W. H. Okamura, M. N. Mitra, M. R. Pirio, S. C. Carey and A. W. Norman, submitted for publication; (d) M. F. Holick, M. Garabedian, H. K. Schnoes and H. F. DeLuca, J. Biol. Chem., 250, 226 (1975).
7. M. J. Bishop and I. Fleming, J. Chem. Soc. (C), 1712 (1969) and references cited.
8. C. Minot, N. T. Anh and L. Salem, J. Am. Chem. Soc., 98, 2678 (1976).
9. (a) M. E. Rennekamp and M. K. Hoffman, Org. Mass. Spectrom., 10, 1067, 1075 (1975); (b) D. H. Williams and G. Hvistendahl, J. Am. Chem. Soc., 92, 6753, 6755 (1974).
10. H. H. Inhoffen and K. Irmscher, Fortschritte Chem. Org. Naturstoffe, 17, 96 (1959).
11. (a) H. H. Inhoffen, G. Quinkert, H. J. Hess and H. Hirschfeld, Chem. Ber., 90, 2544 (1957); (b) S. A. Bakker, J. Lugtenburg and E. Havinga, Rec. Trav. Chim., 91, 1459 (1972).
12. W. G. Dauben and P. Bauman, Tetrahedron Lett., 505 (1961).
13. Besides refs. 4 and 9, see N. J. Turro, D. C. Neckers, P. A. Leermakers, D. Seldner and P. D'Angelo, J. Am. Chem. Soc., 87, 4097 (1965).
14. W. H. Okamura, M. L. Hammond, A. Rego, A. W. Norman and R. M. Wing, submitted for publication.
15. A side by side comparison of the mass spectra for 1, 2 and 5 were made at 15, 20 and 70 eV. The mass spectral data were obtained with an AEI MS-902 mass spectrometer, using a direct insertion probe at temperatures of 50-60°C above ambient (to avoid thermal reactions<sup>10</sup>). In utilizing metastable ion decompositions as evidence for ion structure identity, there are possible pitfalls. For example, see K. Levsen and F. W. McLafferty, J. Am. Chem. Soc., 96, 139 (1974).
16. R. G. Cooks, J. H. Benyon, R. M. Caprioli and G. R. Lester, "Metastable Ions," Elsevier Scientific Publishing Company, Amsterdam, 1973: (a) pp 183-6; (b) pp 41-2.
17. The spectrum of 8 has been previously published. See T. Suda, R. B. Hallick, H. F. DeLuca and H. K. Schnoes, Biochem., 9, 1654 (1970).
18. These spectra were recorded at 70 eV on a Finnigan 1051C Mass Spectrometer.
19. However, this does not imply that cycloaddition of 1<sup>+</sup> (or 2<sup>+</sup> via 1<sup>+</sup>) to a suprasterol-like structure (e.g., 10) necessarily precedes further fragmentation. Other pathways, e.g., cyclization to structure 11 directly, seem equally well feasible.